



USE PROTOCOL FOR R & F[®] *CAMPYLOBACTER JEJUNI/C. COLI* CHROMOGENIC PLATING MEDIUM

- 1 Prepare the R & F[®] *Campylobacter jejuni/C. coli* Chromogenic Plating Medium according to the instructions provided on the packaging labels. After the plates have been poured, they should be stored in the dark for 48 to 72 hours at room temperature or put under a laminated flow hood with the cover removed for 45 to 60 minutes to dry the surface of the agar. After surface drying, the plates can be placed in Petri plate sleeves (slits added to the sleeves to ensure moisture release) and stored at 2-8°C for up to 45 days.
- 2 Inoculate the plates using spread-planting or streaking for isolation, as preferred. Allow the inoculum to completely soak into the agar before inversion and incubation.
- 3 Incubate the plates inverted at 41-42°C under microaerophilic conditions (5 to 7.5% oxygen, 7.5 to 10% carbon dioxide, and 85% nitrogen). This can be achieved with either the gas tank/vacuum assembly or with gas-generating envelopes in an anaerobic jar. Observe the plates after 48 hours.
- 4 *Campylobacter jejuni* and *Campylobacter coli* colonies appear flat to convex, dark salmon; 1.0 to 2.0 mm in diameter with and without a clear ring. Other colonial types appearing on this plating medium that are not presumptive *C. jejuni/C. coli* strains are clear or white and off white (beige).
- 5 Use standard methods for colony confirmation; or an alternative cost effective protocol that can be performed in <3 hours consisting of:
 - a. Gram stain - slender, spiral curved rods; older culture can be spherical or coccoid bodies
 - b. Wet mount to evaluate motility - corkscrew-like motion
 - c. Oxidase test - Oxidase positive
 - d. Commercial Latex Agglutination tests - To determine the presence of *Campylobacter* sp.
 - e. Hippurate hydrolysis - To separate *C. jejuni* from *C. coli*. *C. jejuni* is positive, but *C. coli* is negative. CAUTION: Some hippurate negative *C. jejuni* strains have been reported but infrequently.